

*Asian Research Journal of Agriculture*

*Volume 17, Issue 3, Page 52-65, 2024; Article no.ARJA.119821 ISSN: 2456-561X*

# **Study on Genetic Variability for Forage Yield and Quality Traits in Forage Sorghum [***Sorghum bicolor* **L. Moench]**

# **Santosh a++\* and P. K. Pandey b#**

*<sup>a</sup> Department of Genetics and Plant Breeding, Dr. K. S. Gill Akal College of Agriculture, Eternal University-Baru Sahib, Himachal Pradesh-173101, India. <sup>b</sup> Department of Genetics and Plant Breeding, College of Agriculture, G. B. Pant University of Agriculture and Technology- Pantnagar, U. S. Nagar-263145, (Uttarakhand), India.*

#### *Authors' contributions*

*This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.*

#### *Article Information*

DOI[: https://doi.org/10.9734/arja/2024/v17i3472](https://doi.org/10.9734/arja/2024/v17i3472)

#### **Open Peer Review History:**

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/119821>

*Original Research Article*

*Received: 05/05/2024 Accepted: 06/07/2024 Published: 08/07/2024*

# **ABSTRACT**

The initial research related to germplasm screening was carried out in the experimental area of Instructional Dairy Farm, GBPUAT Pantnagar during kharif season 2018. The experimental material for this experiment consisted of two hundreds and eighty diverse germplasm lines of sorghum along with six checks *viz*., SSG 59-3, Pant Chari- 5, Pant Chari- 6, CSV-21 F, CSH-22S, and CSV-24SS. The germplasm lines were evaluated in Augmented Block Design for nineteen forage yield related traits and thirteen quality traits. The statistical analysis for genetic variability was done using analysis of variance (ANOVA), heritability (h<sup>2</sup>), genotypic coefficient of variation (GCV), phenotypic coefficient of variation (PCV), and genetic advance (GA). The analysis of variance revealed that the genotypes and checks were showing a highly significant differences for all the traits under study

*++ Assistant Professor- Genetics and Plant Breeding;*

*Cite as: Santosh, and P. K. Pandey. 2024. "Study on Genetic Variability for Forage Yield and Quality Traits in Forage Sorghum [Sorghum Bicolor L. Moench]". Asian Research Journal of Agriculture 17 (3):52-65. https://doi.org/10.9734/arja/2024/v17i3472.*

\_

*<sup>#</sup> Professor- Genetics and Plant Breeding;*

*<sup>\*</sup>Corresponding author: E-mail: santosh.8956@gmail.com;*

indicated existence of inherent genetic differences among genotypes for different characters. Mean performance of forage yield and quality related traits exhibited a wide range of variability. GCV and PCV values were observed low for most of the characters whereas high for anthracnose, zonate leaf spot and shoot fly incidence. The value of genetic advance was observed high for leaf area, green fodder yield per plant, dry fodder yield per plant, hydrocyanic acid content, plant height, anthracnose, and shoot fly incidence were relatively more than other characters. Genetic advance as per cent of mean was observed low for days to maturity, dry matter per cent, in-vitro dry matter disappearance, neutral detergent fiber, acid detergent fiber, cellulose content whereas high for leaf area, flag leaf width, panicle length, panicle width, 1000-grains weight, grain yield per plant, green fodder yield per plant, dry fodder yield per plant, total soluble solids, silica content, anthracnose, zonate leaf spot and shoot fly incidence. Heritability values were found high for all the traits under study. The genotypes bearing the desired values for different genetic variability parameters and mean performance for can be exploited in future breeding programmes for the improving forage genotypes. These genotypes can be used as donor parents in crop improvement programme for improving forage yield and quality related traits along with improved resistance against anthracnose, shooty fly and zonate leaf spot.

#### *Keywords: Sorghum; ANOVA; GCV; PCV; heritability.*

# **1. INTRODUCTION**

**"**Sorghum is one of the most important and widely grown crops in the world having the an area of 41.14 million hectares with the production of about 58.72 million tonnes globally whereas 5.00 million hectare and 4.50 million tonnes grain production in India" [1]. "Sorghum is called guinea-corn, dawa, or sorgho in West Africa, durra in the Sudan, mshelia in Ethiopia and Eritrea, mtama in East Africa, kaffir corn in South Africa, and amabele or mabele in several Southern African countries. It is called jowar, cholam, jonna, and jola in India" [2]. Five primary farmed sorghum races are Bicolor, Guinea, Kafir, Durra, and Caudatum [3]. Food, fodder, feed, fuel, and fiber are produced from forage, grain, and sweet sorghum. Due to its drought resistance, fast growth, and outstanding palatability, the crop is grown in tropical and subtropical climates with varying rainfall, temperature, and soil. On small holdings in stress-prone and marginal semi-arid tropics, most of the crop is cultivated. It is produced in Northern Western India for kharif and summer feed. Sorghum is the best kharif fodder for nutrition, with starch (63.68%), high digestibility (50-60%), dry matter (20-35%), sugars (8-17%), crude protein (7.5-10.0%), calcium (0.53%), phosphorus (0.24%), and crude fiber (30-32%). In addition to carbs, it has more iron (Fe) and vitamin B3 than maize and rice. Sorghum is a staple food in Asia and Africa and an important feed crop in the US, Argentina, Mexico, South Africa, and Australia [4]. Any effective breeding program depends on genetic variation and the interactions among various individuals. Plant potential for increased efficiency is determined

by genetic variability; consequently, their utility for breeding is determined as well, which finally might lead to increased forage production. Genetic variability helps to explain the genetic variations between several populations inside a species or between different species. Analyzing the several genetic parameters such as analysis of variance, heritability, genetic advance helps one to determine genetic variability. Higher heterotic expression in F1 and more genetic diversity in segregating populations are the outcomes of parents with higher genetic variability [5]. Plant breeders can choose genetically different parents for purposeful hybridization with the assistance of precise knowledge on the type and extent of genetic variability [6]. The degree and kind of genetic variability determines the genetic enhancement of yield, particularly in self-pollinated crops [7]. In sorghum breeding, hybridization and subsequent selection are two key techniques. The initial step in a plant breeding program using hybridization is the parents' decision. Genetic diversity between parents is required to produce transgressive segregants [8]. Higher levels of genetic variation between parents are associated with higher levels of heterosis in offspring [7]. In sorghum hybridization programs, one suitable tool for parental choice is genetic variability assessment. In order to maximize genetic recombination and perhaps boost output, crossing nurseries must employ parents who have been carefully selected. Given the foregoing, screening the variability of forage sorghum lines, according to yield and quality variables is necessary to determine their suitability for various breeding programs.

# **2. MATERIALS AND METHODS**

The initial research related to germplasm screening was carried out in the experimental area of Instructional Dairy Farm, Nagla, G.B. Pant University of Agriculture and Technology. Pantnagar, District U. S. Nagar, Uttarakhand during *Kharif*, 2018. The experimental material for this experiment consisted of two hundred and eighty diverse germplasm lines of sorghum along with six checks *viz*., SSG 59-3, Pant Chari- 5, Pant Chari- 6, CSV-21 F, CSH-22S, and CSV-24SS. The germplasm lines were evaluated in Augmented Block Design during *Kharif* season of 2018. The experiment was carried out in an Augmented Block Design [9,10], (Federer 1961), with each block containing 35 test entries and 6 checks which were randomly allocated in 8 blocks. All genotypes were sown on 23rd July 2018 in single row of 5 meter length with a row spacing of 45 cm. All the recommended package of practices for sorghum was followed to raise a healthy crop. The observations were recorded ondays to 50% flowering, days to maturity, number of leaves per plant, number of nodes, plant height (cm), leaf length (cm), leaf width (cm), leaf area (cm<sup>2</sup> ), flag leaf length (cm), flag leaf width (cm), stem girth (cm), inter-nodal length (cm), panicle length (cm), panicle width (cm), leaf:stem ratio, 1000-grains weight (gm), grain yield per plant (gm), green fodder yield per plant (gm), dry fodder yield per plant (gm), foliar diseases zonate leaf spot and anthracnose [11], shoot fly (*Atherigona soccata*) incidence (Dead hearts %), dry matter (%), brix %, HCN content (ppm) (Hogg and Ahlagreen, 1942) and Gilchrist et al. (1967) , protein content (%) (Jeckson, 1973), in-vitro dry matter disappearance (IVDMD) % (Erwin and Ellinston, 1959), neutral detergent fiber [12], acid detergent fiber (%) and cellulose (%)(Van Soest, 1991),acid detergent lignin (%), cellulose (%) and silica (%) [12]. The statistical analysis was performed by Indostat Hyderabad (Windostat Version 9.3).

#### **(A) Analysis of variance and means:**

The analysis of variance for the augmented design was done using the method given by Federer [9] as described by Federer and Ragavarao [10] and Petersen [13].

#### **(B) Estimation of variability:**

$$
CV (\%) = \frac{Standard deviation}{Mean} \times 100
$$

Genotypic coefficient of variation (GCV %) =

$$
\frac{\sigma_{\rm g}}{\overline{\rm X}}\times100
$$

Phenotypic coefficient of variation (PCV %) =

$$
\frac{\sigma_{\rm p}}{\overline{\rm X}}\times100
$$

Environmental coefficients of variation (ECV %) =

$$
\frac{\sigma_{\rm e}}{\overline{\rm X}} \times 100
$$

Where,

 $\sigma_g$  = Genotypic standard deviation

 $\sigma_{p}$  = Phenotypic standard deviation

 $\sigma_e$  = Environmental standard deviation

 $\overline{X}$  = Grand mean

#### **(C) Estimation of Heritability**

The heritability in broad sense  $h^2$  (b) was estimated for each character as the ratio of genotypic variance to phenotypic variance by the formula:

Heritability (%) = 
$$
\frac{\sigma_s^2}{\sigma_p^2} \times 100
$$

Where,  $\sigma^2$ <sub>g</sub> = Genotypic variance  $\sigma_{\rm p}^2$  = Phenotypic variance

#### **(D) Genetic Advance:**

The expected genetic advance under selection for the different characters was estimated as suggested by Allard (1960).

$$
GA = h_b^2 \times \text{opi} \times K
$$

Where,

GA = expected genetic advance

 $h_b^2$  = heritability in broad sense

- $\sigma$ <sup>opi</sup> =phenotypic standard deviation for i<sup>th</sup> character
- $K =$  intensity of selection, the value of which is 2.06 at 5 % (Lush, 1949)

# **3. RESULTS AND DISCUSSION**

# **3.1 Analysis of Variance**

Analysis of variance for Augmented Block Design was conducted for nineteen morphological and thirteen nutritional quality traits at different crop maturity stage for different characters Analysis of variance for different yield and quality parameters has been presented in Tables 1. The genotypes and checks were showing highly significant difference for all the traits viz., days to flowering, days to maturity, number of leaves, number of nodes, plant height. Leaf length, leaf width, leaf area, flag leaf length, flag leaf width, stem girth, inter-nodal length, panicle length, panicle width, leaf:stem ratio, 1000-grains weight, grain yield per plant, green fodder yield per plant, dry fodder yield per plant, dry matter percent, total soluble solids, hydrocyanic acid content, protein percent, in-vitro dry matter disappearance, neutral detergent fiber, acid detergent fiber, cellulose content, lignin content, silica content, hemicelluloses content, anthracnose, zonate leaf spot and shoot fly incidence. The blocks were also found highly significant for all the characters under study. This type of result indicated existence of inherent genetic differences among genotypes for different characters. The analysis of variance revealed significant difference among the genotypes which validated further on the basis of genetic and statistical analysis of the data. It revealed that mean squares due to genotypes were found to be significant for all the characters. The findings of present study were found similar with the findings of Desai et al*.* [14], Kadam et al*.* [15], Kumar et al*.* [16], Agrawal et al*.* [17] , Bello et al*.* [18], Rani et al*.* [19], and Jadhav et al. [20], Jain and Patel [21], Kumar (2014), Malik et al*.,* [22], Supriya et al. [23], Abraha et al*.* [24], Arvinth et al*.* [25], Ahlawat et al. [26], Dev et al*.* [27], Dhutmal et al*.* [28], Kavya et al*.* [29], Khandelwal et al*.* [30], Kolekear et al*.* [31], Kumar et al*.* [16], Mofokeng et al*.* [32], Navya et al*.* [33], Nirosh et al*.* [34], Ranjith et al*.* [35], Santosh and Pandey [36], Sen et al*.* [37], Shivaprasad et al*.* [38], Singh et al*.* [39], Subramanian et al*.* [40], Sumon et al*.* [41], Tirkey et al*.* [42], Yaqoob et al*.* [43] and Zinzala et al*.* [44].

# **3.2 Mean Performance and Range of Variation**

The mean performance of 280 genotypes and 6 checks along with their general mean and range values were calculated. General mean, range of variability and best top 10 germplasm are mentioned in Table 2. The range of mean performance for days to flowering ranged from 33 days in EP-124 to 88 days in GP- 2011-110-1 with a general mean of 63.57 days. The range of mean performance for days to maturity ranged from 97 days in R-77 (09R-AGR-26) to 148 days in IS 14357 with a general mean of 127.33 days.

The range of mean performance for number of leaves per plant ranged from 7.30 in CSV-17 to 23.30 in IS 28313 with a general mean of 15.35. The range of mean performance for number of nodes ranged from 5.54 in CSV-17 to 21.32 in IS 28313 with a general mean of 14.13.The range of mean performance for plant height ranged from 138.22 cm in CSV-17 to 528.33 in SMC 8 with a general mean of 362.34. The range of mean performance for leaf length ranged from 56.30 cm in C-43 to 107.20 in IS 12735 with a general mean of 82.01 cm. The range of mean performance for leaf width ranged from 3.22 cm in CO (FS) 29 to 10.52 cm In RS 673 with a general mean of 7.36 cm. The range of mean performance for leaf area ranged from 150.45 cm<sup>2</sup> in IS 23992 to 731.43 cm<sup>2</sup> in SPV 1749 with a general mean of 431.22 cm<sup>2</sup>. The range of mean performance for flag leaf length ranged from 28.15 cm in C-43 to 53.60 in IS 12735 with a general mean of 40.98 cm. The range of mean performance for flag leaf width ranged from 1.61 cm in CO (FS)-29 to 8.31 in CSV 19 with a general mean of 3.73 cm. The range of mean performance for stem girth ranged from 1.23 cm in GMS 1338 to 3.43 cm in SMC 14 with a general mean of 2.41 cm. The range of mean performance for inter-nodal length ranged from 16.50 cm in EJN-58 to 40.23 cm in IS 6045 with a general mean of 27.52 cm. The range of mean performance for panicle length ranged from 5.23 cm in EJN-52 to 42.53 cm in SSG-223 with a general mean of 21.90 cm. The range of mean performance for panicle width ranged from 0.13 cm in PC-1002 to 30.23 cm in SSG-227 with a general mean of 10.55 cm. The range of mean performance for leaf: stem ratio ranged from 0.18 in SPV-1750 to 0.48 in RAJ9-1 with a general mean of 0.29.The range of mean performance for 1000-grains weight ranged from 6.10 gm in CO (FS) 29 to 43.46 gm in UP Chari-1 with a general mean of 21.44 gm. The range of mean performance for grain yield per plant ranged from 24.40 gm in CO (FS) 29 to 165.20 gm in UP Chari -1 with a general mean of 85.04 gm. The range of mean performance for green fodder yield per plant ranged from 80.32 gm in EJ-27 to 835.13 gm in Pant Chari-3 with a general mean of 314.76 gm. The range of mean performance for dry fodder yield per plant ranged from 30.33 gm in EJ-27 to 491.60 gm in IS 14241 with a general mean of 113.24 gm. The range of mean performance for dry matter percent ranged from 29.33 % in IS 9162 to 41.80 % in CSH 19 with a general mean of 35.50 %.The range of mean performance for total soluble solids percent ranged from 2.13 % in SSG-243 to 16.50 % in IS 14756 with a general mean of 6.64 %.The range of mean performance for hydrocyanic acid content ranged from 34.14 ppm in CSV-14 to 118.14 ppm in EJ-27 with a general mean of 85.98 ppm. The range of mean performance for protein percent ranged from 7.12 % in CSV 19 to 15.57 % in IS 14241 with a general mean of 11.00 %.The range of mean performance for Invitro dry matter disappearance ranged from 41.23 % in EJ-30 to 63.25 % in IS 9162 with a general mean of 54.46 %.The range of mean performance for neutral detergent fiberranged from 51.21 % in IS 1219 to 60.18 % in SSG-223 with a general mean of 55.17 %.The range of mean performance for acid detergent fiberranged from 30.57 % in EJ-42 to 40.77 % in IS 18927 with a general mean of 34.97%.The range of mean performance for cellulose content ranged from 26.68 % in UPFS-36 (Pant Chari-7) to 33.22 % in SMC-14 with a general mean of 28.86 %.The range of mean performance for lignin content ranged from 4.13 % in IS 18008-2 to 8.14 % in IS 2363 with a general mean of 5.17 %.The range of mean performance for silica content ranged from 1.12 % in EJ-42 to 3.48 % in IS 18933 with a general mean of 2.05 %.The range of mean performance for hemicelluloses content ranged from 13.81 % in IS 25733 to 29.21 % in UPFS (Pant Chari-7) with a general mean of 21.31 %.The range of mean performance for anthracnose percent from 1.25% in SEVS- 2 to 79.25% in EJN-47 with a general mean of 23.40 %.The range of mean performance for zonate leaf spot percent from 1.12% in SSG- 245 to 46.25% in IS 15008-1 with a general mean of 7.84 %.The range of mean performance for Shoot fly incidencepercent from 2.25% in IS 1478 to 50.75% in R-72 (09R-AGR-23) with a general mean of 20.24 %.

# **3.3 Estimates of Genetic Variability Parameters**

Robinson et al*.* [45] classified heritability values as high (>60%), moderate (30-60%) and low (less than 30 %). Deshmukh et al*.* [46] classified PCV and GCV values as low (0-10%), moderate (10-20%) and high (20% and above). Falconer and Mackay [47] classified genetic advance as per cent of mean as low (0-10%), moderate (10- 20%) and high (20% and above).The estimates of phenotypic coefficient of variation (PCV), genotypic coefficient of variation, genetic advance, genetic advance as percent of mean and heritability (*h 2* ) are presented in Table 3 and Fig. 1. The heritability ranged from 53.32 (cellulose content) to 89.27 (green fodder yield

per plant). Genetic advance ranged from 0.05 (leaf: stem ratio) to 108.11 (leaf area.Genetic advance as percent of mean ranged from 2.70 (neutral detergent fiber) to 72.28 (zonate leaf spot). Phenotypic coefficient of variation ranged from 1.55 (neutral detergent fiber) to 35.12 (zonate leaf spot).Genotypic coefficient of variation ranged from 1.31 (neutral detergent fiber) to 35.09 (zonate leaf spot).

# **3.3.1 Phenotypic coefficient of variation**

Phenotypic coefficient of variation was observed low for days to flowering, days to maturity, number of leaves, number of nodes, plant height, Leaf length, leaf width, flag leaf length, stem girth, inter-nodal length, leaf: stem ratio, hydrocyanic acid content, protein percent, in-vitro dry matter disappearance, neutral detergent fiber, acid detergent fiber, cellulose content, lignin content, hemicelluloses content, moderate for leaf area, flag leaf width, panicle length, panicle width, 1000-grains weight, grain yield per plant, green fodder yield per plant, dry fodder yield per plant, dry matter percent, total soluble solids and silica content whereas high for anthracnose, zonate leaf spot and shoot fly incidence.

Any breeding programme's success is determined on the extent of genetic variability present in the breeding population. The assessment of variability in a crop is critical for identifying lines that can generate further variability, allowing for the artificial selection of desirable, diversified genotypes. Some of the most useful mutations would go unutilized if the breeder had not detected them throughout the selection process.

# **3.3.2 Genotypic coefficient of variation**

Genotypic coefficient of variation was observed low for days to flowering, days to maturity, number of leaves, number of nodes, plant height, Leaf length, leaf width, flag leaf length, stem girth, inter-nodal length, leaf: stem ratio, hydrocyanic acid content, protein percent, in-vitro dry matter disappearance, neutral detergent fiber, acid detergent fiber, cellulose content, lignin content, hemicelluloses content, moderate for leaf area, flag leaf width, panicle length, panicle width, 1000-grains weight, grain yield per plant, green fodder yield per plant, dry fodder yield per plant, dry matter percent, total soluble solids and silica content whereas high for anthracnose, zonate leaf spot and shoot fly incidence.

**120 100 80**  $h 2(\%)$  $\blacksquare$  G.A. **60** G.A.M.  $\blacksquare$  PCV (%) **40**  $\blacksquare$  GCV  $(\% )$ **20**  $\mathcal{L}_{\mathcal{P}}$ 

*Santosh and Pandey; Asian Res. J. Agric., vol. 17, no. 3, pp. 52-65, 2024; Article no.ARJA.119821*

**Fig. 1. Genetic variability, heritability and genetic advance, genotypic and phenotypic coefficients of variation for different characters**

**Table 1. Analysis of variance of augmented block design for different characters in sorghum.**

Source of variation	df	DF	DΜ	NL.	<b>NN</b>	PН		LW	LA.	FLL	FLW	SG
<b>Block (ignoring treatments)</b>		285.531**	247.122**	37.693**	$14.555***$	9865.697**	136.789**	2.883**	20954.890**	42.773**	$0.540**$	0.699**
<b>Treatment (eliminating block)</b>	285	124.529**	126.304**	$5.719**$	$5.739**$	3279.075**	134.666**	2.925**	22583.380**	35.081**	$2.095**$	$0.165**$
Checks		878.283**	711.733**	8.086**	$6.719**$	25290.720*	134.743**	$31.130**$	461307.500**	410.930**	45.798**	$0.462**$
Checks+Var. vs. Var.	280	1.069** 111	115.850*	5.677**	$5.721**$	2886.010**	116.808**	$2.422**$	14749.020**	28.369**	1.314**	$0.159**$
Error	35	0.493	0.857	0.001	0.870	19.110	4.760	0.380	526,770	1.010	0.290	0.010
<b>Block (Eliminating Check+Var.)</b>		$5.036**$	4.286**	$0.606**$	$0.763**$	40.250**	13.329**	$1.011**$	63.785**	$8.197**$	$0.508**$	$0.005**$
<b>Entries (ignoring blocks)</b>	285	131.418**	132.268**	$6.630**$	$6.078**$	3520.402**	137.698**	2.971**	23096.496*	35.930**	$2.096**$	$0.182**$
Varieties	279	115.548**	114.107**	$5.437**$	4.684**	3030.236**	115.917**	$0.959**$	11343.750*	29.138**	$0.561**$	$0.177**$
<b>Checks vs. Varieties</b>		824.934**	2301.952*	332.046*	391.619**	31425.020**	1229.541**	144.613**	1111057**	55.863**	211.695*	$0.047**$

**Continued…….**



**Continued…….**



*\* Significant at 5%, \*\* Significant at 1%.*

*DF= Days to 50% flowering, DM= Days to maturity, NL= Number of leaves per plant, NN= Number of nodes, PH= Plant height (cm), LL= Leaf length (cm), LW= Leaf width (cm), LA= Leaf area (cm<sup>2</sup> ), FLL= Flag leaf length (cm), FLW= Flag leaf width (cm), SG= Stem girth (cm), INL= Internodal length (cm), PL= Panicle length (cm), PW= Panicle width (cm), L:S= Leaf:stem ratio, TGW= 1000-grains weight (gm), GYP=*  Grain yield per plant (gm), GFY= Green fodder yield per plant (gm), DFY= Dry fodder yield per plant (gm), DM= Dry matter (%), TSS= Total soluble solids (%), HCN= HCN content (ppm), PP= Protein content *(%), IVDMD= Iin-vitro dry matter disappearance, NDF= Neutral detergent fiber, ADF=Acid detergent fiber (%), C= Cellulose (%), L=Lignin (%), S= Silica (%), HC= Hemicellulose, A= Anthracnose (%), ZLS= Zonate Leaf Spot, SFI= Shoot Fly Index (%)*

### **Table 2. General mean, range of variability and best top 10 germplasm**





<b>Variables</b>	$h^{2}(\%)$	<b>G.A.</b>	<b>G.A.M.</b>	<b>PCV (%)</b>	GCV (%)
DF	86.60	8.11	12.76	6.21	6.19
DM	84.32	8.16	6.41	3.12	3.11
<b>NL</b>	81.95	1.74	11.34	5.51	5.51
<b>NN</b>	84.82	1.60	11.34	5.98	5.51
<b>PH</b>	82.56	41.59	11.48	5.59	5.57
LL.	86.45	8.29	10.11	5.00	4.91
LW	86.94	1.16	15.75	8.20	7.65
LA	87.66	108.11	25.07	12.31	12.17
<b>FLL</b>	87.14	4.25	10.38	5.11	5.04
<b>FLW</b>	86.40	0.98	26.33	13.75	12.78
SG	86.24	0.29	12.05	5.96	5.85
<b>INL</b>	83.69	3.73	13.54	6.79	6.57
<b>PL</b>	87.62	5.15	23.52	11.56	11.42
<b>PW</b>	87.74	3.94	37.35	18.34	18.13
L:S	88.75	0.05	16.48	8.49	8.00
<b>TGW</b>	87.91	6.16	28.73	14.09	13.95
<b>GYP</b>	88.49	22.07	25.95	12.69	12.60
<b>GFY</b>	89.27	80.68	25.63	12.49	12.44
<b>DFY</b>	88.56	31.72	28.01	13.63	13.60
<b>DM %</b>	85.84	2.47	6.97	3.65	3.38
TSS %	84.25	2.45	36.84	18.15	17.88
<b>HCN</b>	85.94	12.88	14.99	7.35	7.27
<b>PP</b>	85.87	1.71	15.56	7.71	7.55
<b>IVDMD</b>	53.98	2.43	4.47	2.95	2.17
<b>NDF (%)</b>	71.66	1.49	2.70	1.55	1.31
<b>ADF</b>	69.82	1.49	4.25	2.47	2.06
$C\%$	53.32	1.06	3.68	2.44	1.78
L%	82.21	0.65	12.53	6.33	6.08
$S\%$	86.67	0.46	22.53	11.12	10.94
<b>HC %</b>	84.60	2.66	12.49	6.23	6.06
A %	84.52	14.47	61.83	30.02	30.01
ZLS %	85.63	5.67	72.28	35.12	35.09
SFI %	82.56	10.79	53.29	25.98	25.87

**Table 3. Genetic variability, heritability and genetic advance, genotypic and phenotypic coefficients of variation for different characters.**

*DF= Days to 50% flowering, DM= Days to maturity, NL= Number of leaves per plant, NN= Number of nodes, PH= Plant height (cm), LL= Leaf length (cm), LW= Leaf width (cm), LA= Leaf area (cm<sup>2</sup> ), FLL= Flag leaf length (cm), FLW= Flag leaf width (cm), SG= Stem girth (cm), INL= Internodal length (cm), PL= Panicle length (cm), PW= Panicle width (cm), L:S= Leaf:stem ratio, TGW= 1000-grains weight (gm), GYP=*  Grain yield per plant (gm), GFY= Green fodder yield per plant (gm), DFY= Dry fodder yield per plant (gm), DM= Dry matter (%), TSS= Total soluble solids (%), HCN= HCN content (ppm), PP= Protein content *(%), IVDMD= Iin-vitro dry matter disappearance, NDF= Neutral detergent fiber, ADF=Acid detergent fiber (%), C= Cellulose (%), L=Lignin (%), S= Silica (%), HC= Hemicellulose, A= Anthracnose (%), ZLS= Zonate Leaf Spot, SFI= Shoot Fly Index (%)*

## **3.3.3 Genetic advance**

The value of genetic advance was observed high for leaf area, green fodder yield per plant, dry fodder yield per plant, hydrocyanic acid content, plant height, anthracnose, shoot fly incidence were relatively more than other characters such as days to flowering, days to maturity, , Leaf length, leaf width, flag leaf length, flag leaf width, stem girth, inter-nodal length, stem girth, internodal length, panicle length, panicle width, leaf:stem ratio, 1000-grains weight, dry matter percent, total soluble solids, protein percent, invitro dry matter disappearance, detergent fiber, acid detergent fiber, cellulose content, lignin content, silica content and zonate leaf spot.

#### **3.3.4 Genetic advance as percent of mean**

Genetic advance as percent of mean was observed low for days to maturity, dry matter percent, in-vitro dry matter disappearance, neutral detergent fiber, acid detergent fiber, cellulose content moderate for days to flowering, number of leaves, number of nodes, plant height, Leaf length, leaf width, flag leaf length, stem girth, inter-nodal length, leaf:stem ratio, hydrocyanic acid content, protein percent, lignin content, hemicelluloses content whereas high for leaf area, flag leaf width, panicle length, panicle width, 1000-grains weight, grain yield per plant, green fodder yield per plant, dry fodder yield per plant, total soluble solids, silica content, anthracnose, zonate leaf spot and shoot fly incidence.

# **3.3.4 Heritability (***h 2* **)**

Heritability (*h 2* ) was observed high for days to flowering, days to maturity, number of leaves, number of nodes, plant height. Leaf length, leaf width, leaf area, flag leaf length, flag leaf width, stem girth, inter-nodal length, panicle length, panicle width, leaf: stem ratio, 1000-grains weight, grain yield per plant, green fodder yield per plant, dry fodder yield per plant, dry matter percent, total soluble solids, hydrocyanic acid content, protein percent, in-vitro dry matter disappearance, neutral detergent fiber, acid detergent fiber, lignin content, silica content, hemicelluloses content, anthracnose, zonate leaf spot and shoot fly incidence except cellulose content. Heritability and genetic advance are important selection parameters. The estimate of genetic advance is more useful as a selection tool when coupled with heritability estimates [48]. High heritability coupled with high genetic advance was observed for leaf area, flag leaf

width, panicle length, panicle width, 1000- grains weight, grain yield per plant, green fodder yield, dry fodder yield, total soluble solids, silica content, anthracnose, zonate leaf spot and shoot fly incidence. The estimates of genetic advance help in understanding the type of gene action involved in the expression of various yield and quality characters. High values of genetic advance are indicative of additive gene action whereas low values are indicative of non-additive gene action [49]. The findings of present study on genetic variability parameters were observed similar with the findings of Jain and Patel [21], Kumar (2014), Malik et al*.* [22], Supriya et al. [23], Abraha et al*.,* [24], Arvinth et al*.* [25], Ahlawat et al. [26], Dev et al*.* [27], Dhutmal et al*.* [28], Kavya et al*.* [29], Khandelwal et al*.* [30], Kolekear et al. [31], Sheoran et al. [50], Kumar et al*.* [16], Mofokeng et al*.* [32], Navya et al. [33], Nirosh et al. [34], Ranjith et al*.* [35], Santosh and Pandey [36], Sen et al*.* [37], Shivaprasad et al*.*  [38], Singh et al*.* [39], Subramanian et al*.* [40], Sumon et al*.* [41], Tirkey et al*.* [42], Yaqoob et al*.*  [43], and Zinzala et al*.* [44].

#### **. 4. SUMMARY AND CONCLUSION**

Significant differences in the genotypes for every variable under investigation were found by the analysis of variance, and these findings were further supported by the genetic and statistical analyses of the data. It was discovered that mean squares resulting from genotypes were significant for every character. There was a noticeable variation in the average performance across several germplasm lines. For various yield and quality parameters, the range of heritability, GCV, PCV, and genetic advance as a percentage of mean was found to be from low to high. In a program to improve crops, germplasm lines with the appropriate values of variability parameters and mean performance for yield and quality traits can be used. Overall, the results of this study showed that the genotypes under investigation exhibited significant genetic variability. Therefore, in a crop improvement program, there is a chance to directly select superior germplasm for various yield-contributing and quality traits.

# **DISCLAIMER (ARTIFICIAL INTELLIGENCE)**

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

### **REFERENCES**

- 1. USDA Foreign agriculture services; 2019. Available: URL: www.fas.usda.gov.
- 2. Santosh ST, Pandey PK, Rawat RS. Study on genetic variability, heritability and genetic advance for quantitative and quality traits in forage sorghum [*Sorghum bicolor* (L.) Moench. The Pharma Innovation Journal. 2020;9(12):396-401
- 3. De Wet JMJ, Harlan JR. Origin and evolution of guinea sorghums. East Africa Agriculture Journal. 1972;38:114119.
- 4. Miller FR, Kebede Y. Genetic contributions to yield gains in sorghum, 1950 to 1980. In Fehr W.R., ed, Genetic Contributions to Yield in Five Major Crop Plants. Crop Science Society of America Special Publication 7. Crop Science Society of America. Madison, WI, 1984;1-14.
- 5. Shekhawat US, Vijay P, Singhania DL. Genetic divergence in barley (*Hordeum vulgare* L.). Indian J Agric. Res. 2001; 35(2):121-123.
- 6. Arunachalam VA. Genetic distances in plant breeding. Indian J Genet. 1981;4: 226-236
- 7. Joshi AB, Dhawan NL. Genetic improvement of yield with special reference to self-fertilizing crops. Ind J. Genet.and Plant Breed. 1966;26:101-113.
- 8. Joshi BK, Mudwari A, Bhatta MR, Ferrara GO. Genetic diversity in Nepalese wheat cultivars based on agro-morphological traits and coefficients of parentage. Nep Agric Res J. 2004;5:7-17.
- 9. Federer WF. Experimental design. LWW; 1956 Apr 1.
- 10. Federer WT, Raghavarao D. On augmented designs. Biometrics. 1975 Mar 1;29-35.
- 11. Thakur VK, Thakur MK, Gupta RK. Graft copolymers from cellulose: synthesis, characterization and evaluation. Carbohydrate polymers. 2013 Aug 14;97(1):18-25.
- 12. Van Soest PV, Robertson JB, Lewis BA. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. Journal of dairy science. 1991 Oct 1;74(10):3583-97.
- 13. Petersen T. A comment on presenting results from logit and probit models.

American Sociological Review. 1985 Feb 1;50(1).

- 14. Desai SA, Shrotria PK, Singh R. Variability and heterosis for forage yield and its components in inter-specific crosses of forage sorghum. Karnataka J Agric. Sci. 2000;13(2):315-320.
- 15. Kadam DE, Patil FB, Bhor TJ, Harner PN. Line x tester analysis in sweet sorghum hybrids. J Maharatra Agric. Univ. 2001; 25(3):318-319.
- 16. Kumar R, Singh SK, Singh S, Kumar A and Kumar M. Study of genetic variability and heritability of yield and its components in forage sorghum [*Sorghum bicolor* (L.) Moench]. J. pharmacogn. phytochem. 2020;9(5):3103-3106.
- 17. Agrawal M, Shrotria PK. Combining ability analysis for forage yield and quality characters in sorghum [*Sorghum bicolor* (L). Moench]. Forage Research. Agric. Exp. Sta. Res. Bull. 2005;31(1):8-11.
- 18. Bello D, Kadams AM, Simon SY, Mashi DS. Studies on genetic variability in cultivated sorghum [*Sorghum bicolor* (L.) Moench] cultivars of Adamawa State Nigeria. American Eurasian Journal of Agricultural and Environmental Scienc. 2007;2(3):297-302.
- 19. Rani KJ, Rana BS, Swarnalata Kaul, Rao SS, Ganesh M. Parental lines improvement for morphophysiological characters in rabi sorghum. Journal of Maharashtra Agricultural Universities. 2009;34(2):138-140.
- 20. Jadhav RR, Deshmukh DT. Heterosis and Combining Ability Studies in Sorghum (*Sorghum bicolor* (L.) Moench) Over the Environments. Int. J. Curr. Microbiol. App. Sci. 2017; 6(10):3058-3064.
- 21. Jain SK, Patel PR. Genetic parameters and character association in single cut sorghum [*Sorghum bicolor* (L.) Moench]. Research and Review: Journal of agriculture Allied Science; 2013.
- 22. Malik A, Singh SK, Chand P, Singh B, Singh DK. Genetic variability, heritability and genetic advance studies on forage sorghum, Prog. Agric. 2015;15(1):92-94.
- 23. Supriya RG, Deshmukh DT, Sheikh MA, Ghorade RB. Sakhare SB. Genetic studies on forage sorghum [*Sorghum bicolor* (L.) Moench]. Bioinfolet. 2015;12(4B):958-962.
- 24. Abraha T, Githiri MS, Kasili R, Araia W, Nyende BA. Genetic variation among sorghum [*Sorghum bicolor* (L.) Moench] landraces from Eritrea under postflowering

drought stress conditions. American J. of Plant Sci. 2015;6:1410- 1424.

- 25. Arvinth S, Patel RN, Gami RA, Joshi A. Genetic variability, character association and path analysis of forage sorghum [*Sorghum bicolor* (L.) Moench]. Forage Research. 2021;47(2):153-158.
- 26. Ahlawat NK, Arya VK, Kumar P, Singh SK. Genetic divergence in forage sorghum [*Sorghum bicolor* (L.) Moench]. Journal of Applied and Natural Sciences. 2018; 10(1):439-444.
- 27. Dev A, Singh SK, Chand P, Kumar M, Poonia M, Srivastava M. Genetic variability, character association and path analysis in forage sorghum. Journal of Pharmacognosy and Phytochemistry. 2019;8(5):1135-1139.
- 28. Dhutmal RR, More AW, Bhakad KR. Variability studies in kharif sorghum. Journal of Pharmacognosy and Phytochemistry. 2020;9(6):1518-1521.
- 29. Kavya P, Satyanarayana Rao V, Ramana JV, Sreekanth B, Radhakrishna Y and Naffez Umar SK. Variability and divergence studies in sweet sorghum [*Sorghum bicolor* (L.) Moench] for ethanol and its related traits. J. Pharm. Innov. 2021;10(10):1711-1718.
- 30. Khandelwal V, Shukla M, Jodha BS, Nathawat VS, Dashora SK. Genetic parameters and character association in sorghum [*Sorghum bicolor* (L.) Moench]. Indian J. of Sci. and Tech. 2015;8(22):2-4.
- 31. Kolekar NM, Korekar SL and Narkhede GW. Variability, correlation and path analysis in forage sorghum [*Soghum bicolor* (L.) Moench]. Bioinfolet., 2021;18(1a):40-46.
- 32. Mofokeng MA, Shimelis H, Laing M, Shargie N. Genetic variation, heritability and genetic gain for quantitative traits in South African sorghum genotypes. Aust. J. Crop Sci. 2019;13(1):1.
- 33. Navya Y, Singh PK and Sushma B. Genetic Variability and Diversity Analysis for Yield Attributing and Biochemical Parameters of Sorghum [*Sorghum bicolor* (L.) Moench] genotypes. Indian J. Ecol., 2021;48(3):854-859.
- 34. Nirosh, Papineni Venkata, Yuvaraja A, Thangaraj K, Menaka, C. Genetic variability and association studies in segregating generation of red sorghum [*Sorghum bicolor* (L.) Moench] population. Electron. J. Plant Breed. 2021;12(2):521- 524.
- 35. Ranjith P, Ghorade RB, Kalpande VV, Dange AM. Genetic variability, heritability and genetic advance for grain yield and yield components in sorghum. Int. J. Farm Sci. 2017;7(1):90-93.
- 36. Santosh, Pandey PK. Assessment of genetic variability, heritability and genetic advance for yield and quality traits in forage sorghum [*Sorghum bicolor* (L.) Moench]. The Pharma Innovation Journal. 2020;9(10):182-187.
- 37. Sen Ramraj, Saini RK, Singh SK, Kumar A. Study of genetic variability of fodder yield and it's components in forage sorghum [*Sorghum bicolor* (L.) Moench]. Forage Res., 2019;45(2):156-158.
- 38. Shivaprasad T, Girish G, Badigannavar, Ashok, Muniswamy S, Yogesh LN, Ganapathi TR. Genetic variability, correlation and path coefficient studies in sorghum [*Sorghum bicolor* (L.) Moench] mutants. Electron. J. Plant Breed., 2019;10(4):1383-1389.
- 39. Singh SK, Singh A, Kumar R. Genetic variability, character association and path analysis in forage sorghum. Progressive Agriculture. 2016;16(2):214-218.
- 40. Subramanian A, Nirmal Raj R and Elangovan M. Genetic variability and multivariate analysis in sorghum (Sorghum bicolour) under sodic soil conditions. Electron. J. Plant Breed. 2019;10(4):1405- 1414.
- 41. Sumon T, Kumari P, Pahuja SK, Tokas J, Yashveer S. Identification of dual type sorghum genotypes based on correlation and path coefficient studies, Forage Research. 2021;46(4):302-307.
- 42. Tirkey, Sheetal, Jawale LN, More AW. Genetic variability, correlation and path analysis studies in B parental lines of kharif sorghum [*Sorghum bicolor* (L.) Moench]. J. Pharm. Innov. 2021;10(8):624- 628.
- 43. Yaqoob M, Hussain N, Rashid A. Genetic variation and heritability analysis for yield and morphological traits in sorghum [*Sorghum bicolor* (L.) Moench] genotypes. J. Agric. Res. 2015;53(3):331- 343.
- 44. Zinzala S, Davda BK, Modha KG, Pathak VD. Studies on variation, correlation and path coefficient analysis in sorghum [*Sorghum bicolor* L. Moench]. Int. J. of. Agric. Sci. 2018;10(19):7285-7287.
- 45. Robinson HF, Comstock RE, Harvey PH. Estimates of heritability and the degree of

dominance in corn. Agron. J. 1949;41:353- 359.

- 46. Deshmukh SN, MS Basu, PS Reddy. Genetic variability, character association and path coefficients and quantitative traits in Virginia bunch varieties of groundnut. Indian Journal of Agricultural Sciences. 1986; 56:816-821.
- 47. Falconer DS, Trudy FC Mackay. Introduction to Quantitative Genetics 4th ed; 1996.
- 48. Johnson HW, Robinson HE, Comstock RE. Estimates of genetic and environmental variability in soybean. Agron. J. 1955;47:314-318.
- 49. Singh P. Narayanan SS. Biometrical techniques in plant breeding. New Delhi, India: Kalyani Publishers; 1999.
- 50. Sheoran RK, Govila OP, Singh B. Genetic architecture of yield and yield contributing traits in pearl millet. Annals of Agricultural Research. 2000;21:443-445.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

\_ *© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.*

> *Peer-review history: The peer review history for this paper can be accessed here: <https://www.sdiarticle5.com/review-history/119821>*